

# Package ‘vfprogression’

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**Type** Package

**Title** Visual Field (VF) Progression Analysis and Plotting Methods

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**Description** Realization of published methods to analyze visual field (VF) progression. Introduction to the plotting methods (designed by author TE) for VF output visualization. A sample dataset for two eyes, each with 10 follow-ups is included. The VF analysis methods could be found in -- Musch et al. (1999) <[doi:10.1016/S0161-6420\(99\)90147-1](https://doi.org/10.1016/S0161-6420(99)90147-1)>, Nouri-Mahdavi et at. (2012) <[doi:10.1167/iovs.11-9021](https://doi.org/10.1167/iovs.11-9021)>, Schell et at. (2014) <[doi:10.1016/j.ophtha.2014.02.021](https://doi.org/10.1016/j.ophtha.2014.02.021)>, Aptel et al. (2015) <[doi:10.1111/aos.12788](https://doi.org/10.1111/aos.12788)>.

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**plotComponentMatrix**    *General plotting function for multiple 24-2 or 30-2 visual field measurements together:*

**Description**

`plotComponentMatrix` plots the following 24-2 or 30-2 visual field measurement: sensitivity, TD, TD prob, PD, and PD prob:

**Usage**

```
plotComponentMatrix(componentmatrix, ncomp = ncol(componentmatrix),
  plot.ncols = 5, plot.nrows = NULL,
  plot.annot.topleft.function = toString,
  plot.annot.bottomleft.function = function(i) NULL,
  globaltitle = sprintf("k = %i", ncol(componentmatrix)),
  globalannotright = NULL,
  zmin = -ceiling(max(abs(c(min(componentmatrix),
    max(componentmatrix))))), zmax = -zmin,
  color.pal = colorRampPalette(c("red", "white", "blue"), space =
  "Lab")(256), td.probabilities = FALSE,
  show.colorbar = !td.probabilities, titleheight = 0.2, ...)
```

**Arguments**

- componentmatrix**    a matrix or data frame, column represents different eyes and rows are the VF measurements of the same type (sensitivity, TD, TD prob, PD, or PD prob).
- ncomp**    a numeric variable defines the number of components to be plotted (default: all).
- plot.ncols**    a numeric variable defines the number of columns to be plotted (default: 5).
- plot.nrows**    a numeric variable defines the number of rows to be plotted (default: NULL (automatically calculated)).
- plot.annot.topleft.function**    a function(i) that is given to any subplot i to create its top left annotation.

```

plot.annot.bottomleft.function
  a function(i) that is given to any subplot i to create its bottom left annotation
  (default: returns NULL).

globaltitle    a string for global title (default: k = ncomp; set to NULL to suppress global
               title).

globalannotright
  a string annotation to the right of the global title (default: NULL).

zmin          minimum value of the color scale (default: auto defined).

zmax          maximum value of the color scale (default: auto defined).

color.pal      an object that defines color scale theme (default: colorRampPalette(c("red",
               "white", "blue"), space = "Lab")(256)).

td.probabilities
  a logic variable indicates whether to plot TD probability symbols instead of TD
  colors (default: FALSE).

show.colorbar   a logic variable indicates whether to show a global colorbar (default: !td.probabilities).

titleheight    a numeric variable defines the height of the title relative to height of row one.

...
  other variables to be added.

```

## Value

heatmap for sensitivity, TD and PD input. Value plot for TD prob and PD prob input.

## Examples

```

data(vfseries)
componentmatrix = t(vfseries[1:10, grep('^s[0-9]+', colnames(vfseries))])
globaltitle = paste("Sensitivities, k = ", ncol(componentmatrix), sep = '')
plotComponentMatrix(componentmatrix, globaltitle = globaltitle)
componentmatrix = t(vfseries[1:10, grep('^td[0-9]+', colnames(vfseries))])
globaltitle = paste("TDs, k = ", ncol(componentmatrix), sep = '')
plotComponentMatrix(componentmatrix, globaltitle = globaltitle)
componentmatrix = t(vfseries[1:10, grep('^pd[0-9]+', colnames(vfseries))])
globaltitle = paste("PDs, k = ", ncol(componentmatrix), sep = '')
plotComponentMatrix(componentmatrix, globaltitle = globaltitle)
componentmatrix = t(vfseries[1:10, grep('^tdp[0-9]+', colnames(vfseries))])
globaltitle = paste("TD Probs, k = ", ncol(componentmatrix), sep = '')
plotComponentMatrix(componentmatrix, globaltitle = globaltitle, td.probabilities = TRUE)
componentmatrix = t(vfseries[1:10, grep('^pdp[0-9]+', colnames(vfseries))])
globaltitle = paste("PD Probs, k = ", ncol(componentmatrix), sep = '')
plotComponentMatrix(componentmatrix, globaltitle = globaltitle, td.probabilities = TRUE)

```

---

**plotfield.normalized** *Single plotting function for one 24-2 or 30-2 visual field measurement:*

---

## Description

**plotfield.normalized** plots the following 24-2 or 30-2 visual field measurement: sensitivity, TD, TD prob, PD, and PD prob:

## Usage

```
plotfield.normalized(eigenfields, component = 1,
zmin = -max(abs(c(min(eigenfields), max(eigenfields)))), 
zmax = max(abs(c(min(eigenfields), max(eigenfields)))), 
color.pal = colorRampPalette(c("red", "white", "blue"), space =
"Lab")(256), show.colorbar = TRUE, topleftannotation = NULL,
bottomleftannotation = NULL, labelcex = 2, ...)
```

## Arguments

<b>eigenfields</b>	a vector contains Sensitivity/TD/PD measurement. For 24-2 VF eigenfields should have 52 or 54 elements. For 30-2 VF, eigenfields should have 74 or 76 elements.
<b>component</b>	Number of components to be plotted (default: 1).
<b>zmin</b>	minimum value of the color scale (default: auto defined).
<b>zmax</b>	maximum value of the color scale (default: auto defined).
<b>color.pal</b>	an object that defines color scale theme (default: colorRampPalette(c("red", "white", "blue"), space = "Lab")(256)).
<b>show.colorbar</b>	a logic value to show colorbar (default: TRUE).
<b>topleftannotation</b>	a string annotation shown on the top left side of the plot (default: NULL).
<b>bottomleftannotation</b>	a string annotation shown on the bottom left side of the plot (default: NULL).
<b>labelcex</b>	a numeric variable for label size (default: 2).
<b>...</b>	other variables to be added.

## Value

heatmap for sensitivity, TD and PD input

## Examples

```
data(vfseries)
eigenfields = t(vfseries[1, grepl('^s[0-9]+', colnames(vfseries))])
plotfield.normalized(eigenfields)
title(main = "Sensitivity", line = 3)
```

```

eigenfields = t(vfseries[1, grepl('^td[0-9]+', colnames(vfseries))])
plotfield.normalized(eigenfields)
title(main = "Total Deviation", line = 3)
eigenfields = t(vfseries[1, grepl('^pd[0-9]+', colnames(vfseries))])
plotfield.normalized(eigenfields)
title(main = "Pattern Deviation", line = 3)

```

**plotTdProbabilities** *Value plotting function for 24-2 or 30-2 visual field measurement:*

## Description

**plotTdProbabilities** plots the following 24-2 or 30-2 visual field measurement: TD probs, and PD probs:

## Usage

```
plotTdProbabilities(tdprob, cex = 2, rectangle.color = "black",
rectangle.width = 0.16, margins = c(2, 1, 2, 2) + 0.1, ...)
```

## Arguments

<b>tdprob</b>	a vector contains TD probs/PD probs measurement. For 24-2 VF tdprob should have 52 or 54 elements. For 30-2 VF, tdprob should have 74 or 76 elements.
<b>cex</b>	a numeric variable for label size (default: 2).
<b>rectangle.color</b>	a string variable defines label color (default: 'black').
<b>rectangle.width</b>	a numeric variable defines label width (default: '0.16').
<b>margins</b>	a vector define the plot margins (default: c(2, 1, 2, 2)+0.1).
<b>...</b>	other variables to be added.

## Value

value plot for TD prob and PD prob input.

## Examples

```

data(vfseries)
tdprob = t(vfseries[1, grepl('^tdp[0-9]+', colnames(vfseries))])
plotTdProbabilities(tdprob)
title(main = "Total Deviation Probability", line = 3)
tdprob = t(vfseries[1, grepl('^pdp[0-9]+', colnames(vfseries))])
plotTdProbabilities(tdprob)
title(main = "Pattern Deviation Probability", line = 3)

```

**plotTDvalues***Value plotting function for 24-2 or 30-2 visual field measurement:***Description**

`plotTDvalues` plots the following 24-2 or 30-2 visual field measurement: sensitivity, TD, and PD:

**Usage**

```
plotTDvalues(tds, cex.tds = 1, textcolor = function(x) "black",
             show.lines = T, ...)
```

**Arguments**

- |                         |   |
|-------------------------|---|
| <code>tds</code>        | a vector contains sensitivity/TD/PD measurement. For 24-2 VF tds should have 52 or 54 elements. For 30-2 VF, tds should have 74 or 76 elements. |
| <code>cex.tds</code>    | a numeric variable for label size (default: 1).   |
| <code>textcolor</code>  | a function defines the label color.   |
| <code>show.lines</code> | a logical variable indicates whether to show the horizontal and vertical lines.   |
| <code>...</code>        | other variables to be added.  |

**Value**

value plot for sensitivity, TD and PD input.

**Examples**

```
data(vfseries)
tds = t(vfseries[1, grepl('^s[0-9]+', colnames(vfseries))])
plotTDvalues(tds)
title(main = "Sensitivity", line = 3)
tds = t(vfseries[1, grepl('^td[0-9]+', colnames(vfseries))])
plotTDvalues(tds)
title(main = "Total Dviation", line = 3)
tds = t(vfseries[1, grepl('^pd[0-9]+', colnames(vfseries))])
plotTDvalues(tds)
title(main = "Pattern Dviation", line = 3)
```

---

progression	<i>general progression function</i>
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## Description

progression returns the progression criterion with four methods. plr.nouri.2012, vfi, schell2014, cigts

## Usage

```
progression(vfseries, method = c("plr.nouri.2012", "vfi", "schell2014",
  "cigts"))
```

## Arguments

- vfseries** is a data frame. MUST contain the following columns: 'yearsfollowed', and 'eyeid'. Rows represent the single measurements. Other requirements, such as number of minimum measurements (rows), and necessary VF measurements could be found in each progression method's documentation
- method** selected from one or more from: plr.nouri.2012, vfi, schell2014, cigts. Default it ...

## Value

"stable", "worsening", or "improving" of measurements in `measmatrix`

## See Also

<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4495761/>

## Examples

```
data(vfseries)
progression(vfseries)
progression(vfseries[vfseries$eyeid == 1,])
progression(vfseries[vfseries$eyeid == 2,])
progression(vfseries, method=c("cigts"))
```

`progression.cigts`      *CIGTS VF progression*

## Description

`progression.cigts` returns the progression of visual field test based on 52 or 54 total deviation probabilities (tdp). CIGTS VF progression (Musch et al., 1999).

## Usage

```
progression.cigts(measmatrix)
```

## Arguments

`measmatrix` is a data frame. MUST contain the following columns: 52/54 TD probs (column names MUST be 'tdp1' ~ 'tdp52' or 'tdp1' ~ 'tdp54'), 'yearsfollowed', and 'eyeid'. Rows represent the single measurements. The minimum measurements (rows) is 5.

## Value

"stable", "worsening", or "improving" of measurements in `measmatrix`. Note: If a VF series is temporarily improving and temporarily worsening, it is assumed to be "stable" overall

## References

[http://www.aojournal.org/article/S0161-6420\(99\)90147-1/abstract](http://www.aojournal.org/article/S0161-6420(99)90147-1/abstract)

## Examples

```
data(vf.cigts)
colnames(vf.cigts)
progression.cigts(vf.cigts)
progression.cigts(vf.cigts[vf.cigts$eyeid == 1,])
progression.cigts(vf.cigts[vf.cigts$eyeid == 2,])
```

`progression.plr.nouri.2012`

*Nouri-Mahdavi 2012 VF progression*

## Description

`progression.plr.nouri.2012` returns the progression criterion, using Pointwise Linear Regression (PLR) progression detection method according to Nouri-Mahdavi et al. (2012).

**Usage**

```
progression.plr.nouri.2012(measmatrix)
```

**Arguments**

`measmatrix` is a data frame. MUST contain the following columns: 52/54 TD (column names MUST be 'td1' ~ 'td52' or 'td1' ~ 'td54'), 'yearsfollowed', and 'eyeid'. Rows represent the single measurements. The minimum measurements (rows) is 3.

**Value**

"stable", "worsening", or "improving" of measurements in `measmatrix`

**See Also**

<https://www.ncbi.nlm.nih.gov/pubmed/22427560/>

**Examples**

```
data(vf.plr.nouri.2012)
colnames(vf.plr.nouri.2012)
progression.plr.nouri.2012(vf.plr.nouri.2012)
progression.plr.nouri.2012(vf.plr.nouri.2012[vf.plr.nouri.2012$eyeid == 1,])
progression.plr.nouri.2012(vf.plr.nouri.2012[vf.plr.nouri.2012$eyeid == 2,])
```

progression.schell2014

*Schell 2014 VF progression*

**Description**

`progression.schell2014` returns the progression criterion after Schell et al. 2014, which is essentially like CIGTS but with MD, and only one follow-up is enough to confirm progression.

**Usage**

```
progression.schell2014(measmatrix)
```

**Arguments**

`measmatrix` is a data frame. MUST contain the following columns: 'md' (mean deviation) and 'eyeid'. Rows represent the single measurements. The minimum measurements (rows) is 4.

**Value**

"stable", "worsening", or "improving" of measurements in `measmatrix`. Note: If a VF series is temporarily improving and temporarily worsening, it is assumed to be "stable" overall

**See Also**

<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC4495761/>

**Examples**

```
data(vf.schell2014)
colnames(vf.schell2014)
progression.schell2014(vf.schell2014)
progression.schell2014(vf.schell2014[vf.schell2014$eyeid == 1,])
progression.schell2014(vf.schell2014[vf.schell2014$eyeid == 2,])
```

**progression.vfi**

*progression according to VFI (significant slope, p<=0.05)*

**Description**

`progression.vfi` returns the progression criterion used in Aptel et al. (2015).

**Usage**

```
progression.vfi(measmatrix)
```

**Arguments**

<code>measmatrix</code>	is a data frame. MUST contain the following columns: 'vfi' (visual field index), 'yearsfollowed', and 'eyeid'. Rows represent the single measurements. The minimum measurements (rows) is 3.
-------------------------	--

**Value**

"stable", "worsening", or "improving" of measurements in timepoints

**See Also**

<https://www.ncbi.nlm.nih.gov/pubmed/26095771/>

**Examples**

```
data(vf.vfi)
colnames(vf.vfi)
progression.vfi(vf.vfi)
progression.vfi(vf.vfi[vf.vfi$eyeid == 1,])
progression.vfi(vf.vfi[vf.vfi$eyeid == 2,])
```

---

**vf.cigts***Combined Visual Field Series for General Progression Method*

---

**Description**

Data

**Usage**

```
data(vf.cigts)
```

**Format**

A data frame sample for CIGTS progression method, which includes visual field related measurement for two eyes, each with 10 follow-ups. Rows represent the single measurements.

**Source**

**eyeid** eyeid, labeled as 1,2... for different eyes.

**yearsfollowed** follow-up years. The minimum measurements /rows for one eye is 5.

**tdp1-tdp54** 52 total deviation probability, or 'tdp' measurements. The minimum measurements, or rows for one eye is 5. ...

**Examples**

```
data(vf.cigts)
colnames(vf.cigts)
progression.cigts(vf.cigts)
progression.cigts(vf.cigts[vf.cigts$eyeid == 1,])
progression.cigts(vf.cigts[vf.cigts$eyeid == 2,])
```

---

**vf.plr.nouri.2012***Combined Visual Field Series for General Progression Method*

---

**Description**

Data

**Usage**

```
data(vf.plr.nouri.2012)
```

**Format**

A data frame sample for Pointwise Linear Regression (PLR) method according to Nouri-Mahdavi 2012 progression, which includes visual field related measurement for two eyes, each with 10 follow-ups. Rows represent the single measurements.

## Source

**eyeid** eyeid, labeled as 1,2... for different eyes  
**yearsfollowed** follow-up years. The minimum measurements, or rows, for one eye is 3  
**td1-td54** 52 total deviation, or 'td' measurements. The minimum measurements, or rows, for one eye is 3 ...

## Examples

```
data(vf.plr.nouri.2012)
colnames(vf.plr.nouri.2012)
progression.plr.nouri.2012(vf.plr.nouri.2012)
progression.plr.nouri.2012(vf.plr.nouri.2012[vf.plr.nouri.2012$eyeid == 1,])
progression.plr.nouri.2012(vf.plr.nouri.2012[vf.plr.nouri.2012$eyeid == 2,])
```

vf.schell2014

*Combined Visual Field Series for General Progression Method*

## Description

Data

## Usage

```
data(vf.schell2014)
```

## Format

A data frame sample for progression method by Schell et al. 2014, which includes visual field related measurement for two eyes, each with 10 follow-ups. Rows represent the single measurements.

## Source

**eyeid** eyeid, labeled as 1,2... for different eyes.  
**md** mean deviation measurements. The minimum measurements, or rows, for one eye is 4. ...

## Examples

```
data(vf.schell2014)
colnames(vf.schell2014)
progression.schell2014(vf.schell2014)
progression.schell2014(vf.schell2014[vf.schell2014$eyeid == 1,])
progression.schell2014(vf.schell2014[vf.schell2014$eyeid == 2,])
```

---

**vf.vfi***Combined Visual Field Series for General Progression Method*

---

**Description**

Data

**Usage**

```
data(vf.vfi)
```

**Format**

A data frame for CIGTS progression example, which includes visual field related measurement for two eyes each with 10 follow-ups.

**Source**

**eyeid** eyeid, labeled as 1,2... for different eye groups.

**yearsfollowed** follow-up years. The minimum measurements, or rows, for one eye is 3.

**vfi** visual field index. The minimum measurements, or rows, for one eye is 3. ...

**Examples**

```
data(vf.vfi)
colnames(vf.vfi)
progression.vfi(vf.vfi)
progression.vfi(vf.vfi[vf.vfi$eyeid == 1,])
progression.vfi(vf.vfi[vf.vfi$eyeid == 2,])
```

---

**vfseries***Combined Visual Field Series for General Progression Method*

---

**Description**

Data

**Usage**

```
data(vfseries)
```

**Format**

A data frame sample including the following visual field related measurement for two eyes, each with 10 follow-ups.

### Source

**eyeid** eyeid, labeled as 1,2... for different eyes.  
**nvisit** number of visits.  
**yearsfollowed** follow-up years.  
**distprev** to be updated.  
**age** in years.  
**righteye** 1 as right eye, 0 as left eye.  
**malfixrate** VF test malfixation rate.  
**ght** glaucoma hemifield test result.  
**vfi** visual field index.  
**md** mean deviation.  
**mdprob** mean deviation probability.  
**psd** pattern standard deviation.  
**psdprob** pattern standard deviation probability.  
**s1-s54** 52 sensitivity measurements.  
**td1-td54** 52 total deviation measurements.  
**tdp1-tdp54** 52 total deviation probability measurements.  
**pdp1-pdp54** 52 pattern deviation probability measurements. ...

### Examples

```

data(vfseries)
progression(vfseries)
progression(vfseries[vfseries$eyeid == 1,])
progression(vfseries[vfseries$eyeid == 2,])
progression(vfseries, method=c("cigts"))
progression.cigts(vfseries)
progression(vfseries, method=c('plr.nouri.2012', 'schell2014', 'vfi'))
  
```

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